

# Antibiogram Development for Neonatal Intensive Care Unit at Referral Healthcare Center in Sana'a City, Yemen

Submitted By

Massoud Khalid Rassam

Aimen Ahmed Qasem Alhebshi

Noman Mohammed Abdulmajeed

Mohammed Hassan Al Nuzaili

Majde Mohammed Saif

Ala'a Fuod Alsalowi

Lool Abdulrahman Amr

Yusra Ahmed Al Mashmali

Ghada Adnan Salah

Amr Abdullah Nasher

Khalid Ali Alodini

Supervised by

Associate. Prof Dr \ Mokhtar Al-Ghorafi

Dr \ Mohammed Abdullah Ali Kubas

A Graduation Research Project Submitted as Fulfilment of The  
Requirement for the Degree of PharmD

Emirates International University

2022





## Acknowledgements

Firstly, we praise God Almighty very much for what we have honored with from the completion of this study, then we thank our family for all support period of 6th years of study, who granted us the gift of their unwavering belief in our ability to accomplish this goal: thanks you for your support and patience, for Emirates International University and for Dean College of Medicine and Health sciences Prof \ Saleh Al-Daheri and for our head of department and our Supervisor Associate. Prof Dr\ Mokhtar Abdulhafiz Al-Ghorafi for all support and encouragement during period of study.

Finally, a special thanks to Dr\ Mohammed Abdullah kubas for his continuous support, encouragement and leadership, and for university science and technology hospital.





## *Table of Contents*

<b>1.1. Acknowledgements</b>	<b>iii</b>
<b>1.2. Table of Contents</b>	<b>iv</b>
<b>1.3. List of Tables</b>	<b>vi</b>
<b>1.4. Abbreviation</b>	<b>viii</b>
<b>1.5. Abstract</b>	<b>ix</b>
<b>2. Chapter One</b>	
<b>2.1. Introduction</b>	<b>13</b>
<b>3. Chapter Two</b>	
<b>3.1. Literature Review</b>	<b>17</b>
<b>3.1.1. Infection in Neonatal</b>	<b>17</b>
<b>3.1.2. Antimicrobial Resistance</b>	<b>18</b>
<b>3.1.3. Resistance in Gram-Negative Infections</b>	<b>18</b>
<b>3.1.4. Resistance in Gram-Positive Infections</b>	<b>19</b>
<b>4. Chapter Three</b>	
<b>4.1. Methodology</b>	<b>22</b>
<b>4.1.1. Patients and Study Site, Study Design</b>	<b>22</b>
<b>4.1.2. Lab Techniques</b>	<b>22</b>
<b>4.1.3. Data Collection</b>	<b>22</b>
<b>4.1.4. Data Analysis</b>	<b>22</b>
<b>4.1.5. Antibigram Development</b>	<b>22</b>
<b>4.1.6. Ethics Statement</b>	<b>23</b>
<b>5. Chapter Four</b>	
<b>5.1.1. Result</b>	<b>25</b>
<b>5.1.2. Isolated And Number of Bacterial Species That Included in The Antibigram</b>	<b>26</b>
<b>5.1.3. Isolated of Gram Positive Of Bacterial Species</b>	<b>26</b>





6. Chapter Five	
6.1.1.	
<i>Discussion</i>	35
6.1.2. Limitation	37
7. Chapter Six	
7.1.1.	
<i>Conclusions</i>	39
7.1.2.	
<i>Recommendation</i>	39
8. Appendices	42
8.1.1.	
<i>Appendix A Pervious Work</i>	44
8.1.2.	
<i>Appendix B Data Collected</i>	45
8.1.3.	
<i>Appendix C Data Analysis</i>	48
8.1.4	
<i>Appendix D Premission Letter of EIU</i>	48





## List of Tables

<i>Table 1 Gender f Isolated</i>	25
<i>Table 2 The source of isolations with their numbers and frequency.</i>	25
<i>Table 3 The type and number of bacteria isolated</i>	26
<i>Table 4 The type and number of bacteria isolated</i>	26
<i>Table 5 Staphylococcus, coagulase negative sensitivety</i>	27
<i>Table 6 Streptococcus sp sensitivity</i>	28
<i>Table 7 Straphylococcus aureus sensitivty</i>	28
<i>Table 8 Enterococcus sp. sensitive</i>	29
<i>Table 9 Staphylococcus hominis sinsitive</i>	29
<i>Table 10 Antibigram for gram postive</i>	35
<i>Table 11 Acinetobacter sensitive</i>	31
<i>Table 12 Escherichia coli sensitive</i>	32
<i>Table 13 Enterobacter sp sensitive</i>	32
<i>Table 14 Klebsiella sp sensitive</i>	33
<i>Table 15 Klebsiella pneumonia sensitive</i>	33
<i>Table 16 Pseudomonas sp sensitive</i>	34
<i>Table 17 Burkholderia cepacia sensitive</i>	34
<i>Table 18 Antibigram for gram negative</i>	35





## ABBREVIATION

AB	Antibiotic
AMR	Antimicrobial resistance
BSI	Bloodstream infections
CDC	Centers for Disease Control and Prevention
CoNS	Coagulase negative staphylococci
CPE	Carbapenemase-producing Enterobacteriaceae.
CRE	Carbapenem-Resistant Enterobacteriaceae
E.coli	Escherichia coli
ESBL	Extended spectrum beta lactamases
G +ve	Gram positive
G -ve	Gram negative
GBS	Group B streptococcus
GN	Gram negative
GP	Gram positive
HAIs	Healthcare associated infections
HIV	Human immunodeficiency virus
HSV	Herpes simplex virus
I	Intermediate
ICU	Intensive care unit
ICUHTLV	Human T-lymphotrophic virus type 1
MRSA	Methicillin-resistant Staphylococcus aureus
NICU	Neonates intensive critical care sensitive
No	Number of





P. aeruginosa	Pseudomonas aeruginosa
PBP	Penicillin-Binding Protein
R	Resistance
S	Sensitive
S. aureus	Staphylococcus aureus
SCC	Staphylococcal cassette chromosome
TMP/SMX	Trimethoprim/sulfamethoxazole
USTH	University science and technology hospital
UTIs	Urinary Tract Infections
VRSA	Vancomycin Resistance Staphylococcus Aureus
WHO	World Health Organization





## ABSTRACT

### ***Background***

The neonate immunity is very low and infections among them could lead to life threatening consequences. Antibiotic resistance is considered one of the most challenging area in the treatment of infectious diseases worldwide and could lead to treatment failure among neonate.

### ***Objective***

The overall aim of this study was to develop antibiogram that specific for neonate at USTH, Sana'a, Yemen.

### ***Methodology***

This is retrospective study, the data was collected from the USTH database and WOHNET program was used for analysis and development of USTH antibiogram

### ***Result***

The G +ve was the most common type of bacteria encounter among neonate, followed by G -ve bacteria. From G +ve bacteria, staphylococcus coagulase negative and Streptococcus spp were the most common isolated bacteria, and klebsiella spp and Burkholderia cepacia were the most isolated among G-ve bacteria. The most sensitives antibiotic for G+ve bacteria was vancomycin and linezolid, whereas, carbapenem, colistin, and polymyxin B were the most sensitive antibiotics for G-ve bacteria.

### ***Conclusion***

The resistance for antimicrobial agent was high among neonate patient ether for G+ve positive or G-ve bacteria.





# **Chapter One**

## **Introduction**





## INTRODUCTION

Neonates or newborns can be defined as babies from the time of delivery up to four weeks old.<sup>(1)</sup> Neonates possess an incompetent innate or/and adaptive immunity, which make them more susceptible and less able to compete with infections caused by numerous pathogenic microorganisms.<sup>(2)</sup>

Bacterial infections in newborns can range from mild to severe and life-threatening infections, include Bloodstream infections (BSIs) are the most common. They can occur in isolation or in association with urinary tract infections (UTIs) and meningitis. Endocarditis, osteomyelitis, pyogenic arthritis, ventilator associated pneumonia , peritonitis, conjunctivitis, and skin abscesses are important, less common HAIs<sup>(3)</sup>.

Based on the age of neonates when contracting the infection, the infection can be categorized into two groups: early (during the first 7 days of life) and late (after 7 days from delivery) onset.<sup>(4)</sup>

This is because the immune system of neonates is rapidly developing as they are growing, thus each stage of age may possess different level of potency to fight infections.<sup>(4)</sup> This fact might explain the reasons behind observed variation of causative bacteria according to baby's age group.<sup>(2)</sup> Resistance of bacteria to antibiotics is usually caused by genetic modifications as a result of the irrational use of antibiotics.<sup>(5,6)</sup>

The rise in antimicrobial resistance (AMR) continues to be a global crisis. Collectively, antimicrobial-resistant pathogens caused more than 2.8 million infections and over 35,000 deaths annually from 2012 through 2017, according to the 2019 Centers for Disease Control and Prevention (CDC) Antibiotic Resistance Threats in the United States Report.<sup>(7)</sup>

Multiple factors transcending disciplines contribute to the development of AMR, with inappropriate use of antibiotics regarded as a major contributing factor according to the report by the WHO Global Action Plan on antimicrobial resistance.<sup>(8)</sup> The increase level of resistance against antibiotic drugs used to treat bacterial infections associated with sepsis and UTI in neonates is very alarming worldwide.<sup>(9)</sup>

On national data of ABR reported by the World Health Organization (WHO) indicates *Escherichia coli* and *Klebsiella pneumoniae* resistance to 3rd generation cephalosporin at 2–70% and 8–77%, respectively.<sup>(10)</sup>





*E. coli* resistance to fluoroquinolones is reported at 14–71% [10]. Methicillin-resistant *Staphylococcus aureus* (MRSA) is reported to make up 12–80% of *S. aureus* isolates while *Streptococcus pneumoniae* resistance to penicillin is at 3–16%.<sup>(10)</sup> Available literature from SSA supports this national data. *E. coli* and *K. pneumoniae* generally have low susceptibility to penicillin, cephalosporin, fluoroquinolones, and trimethoprim/sulfamethoxazole (TMP/SMX) while maintaining high susceptibility to carbapenems and amikacin.<sup>(11,12)</sup>

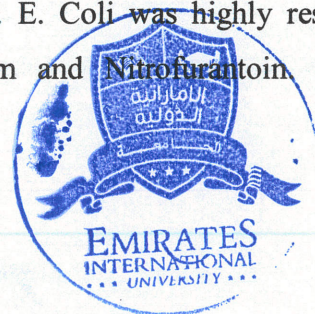
*Klebsiella pneumoniae* is a gram-negative opportunistic bacterium responsible for community- and hospital-acquired infections. The mortality rate of neonatal sepsis caused by *K. pneumoniae* ranges from 18% to 68%.<sup>(13)</sup> On local data reported in Yemen according to USTH in Sana'a period from 2006 to 2013. The most frequently as gram negative isolated species from the inpatients admitted to the departments of the USTH were *E. coli* and *Acinetobacter* species followed by *Klebsiella* species and *P. aeruginosa*.

However, the most frequently isolated Gram-negative bacteria, *Acinetobacter* species has the highest resistance rate to the most commonly used antibiotics, where only polymyxin B is effective against this species. *P. aeruginosa* shows an unchanging rate of resistance to antibiotics in the USTH despite being quite resistant to antibiotics on a global scale.<sup>(14)</sup>

Whereas another study in Aden, Yemen *Staphylococcus* spp. followed by *E. coli*, *Pseudomonas* spp., and *Klebsiella pneumoniae*, were the most widespread pathogenic bacteria in several isolates. Overall bacterial resistance was common for old antibiotics, such as the combination of sulfamethoxazole with trimethoprim, followed by amoxicillin and clavulanate. Additionally, cephalosporin had a relatively higher resistance rate than other antibiotics.

The study also showed moderate bacterial resistance toward gentamycin, azithromycin, cefoxitin, and ciprofloxacin. However, a lower percentage of resistance was present for the combination of ampicillin with sulbactam, ertapenem, and levofloxacin.<sup>(18)</sup>

Pervious antibiogram that was done by clinical (antibiogram) Gram negative organisms *E. Coli* and *Acinetobacter* species were the most isolated gram negative (GN) bacteria, with 246(35.7%) and 148(21.4%) specimens, respectively. *E. Coli* was highly resistant to all antibiotics except Amikacin, Meropenem, Imipenem and Nitrofurantoin. *E. coli* had





susceptibilities 56% to gentamicin and 56% to Piperacillin /Tazobactam. Acinetobacter and klebsiella spp were highly resistant to all antibiotics.

Klebsiella pneumoniae had susceptibilities 59% to Amikacin, 66% to Imipenem and 71% to Meropenem. Pseudomonas aeruginosa had susceptibilities 64%to amikacin, 59%to gentamicin and 58% to Imipenem. Pseudomonas spp was highly resistant to all antibiotics except Imipenem. pseudomonas spp had susceptibilities 55% to Piperacillin /Tazobactam and 59% to Meropenem.

Gram positive organisms, There was 123(43.3%) coagulase negative staphylococci (CONS) isolates, 115(40.5%) S. aureus isolates, 23(8.1%) enterococcus spp isolates, 23(8.1%) streptococcus spp isolates CoNS had decreased susceptibility to Azithromycin(17%), Erythromycin (21%), Norfloxacin(25%) and Ampicillin /Sulbactam(45%) while maintaining susceptibility to Vancomycin, Linezolid, Imipenem, Amikacin. Enterococcus spp was highly resistant to all antibiotics except linezolid and Vancomycin. E. spp had susceptibility to Meropenem (60%).

S. Aureus had highly susceptibility to Imipenem and vancomycin. MRSA had highly resistant to all antibiotics except Vancomycin, linezolid and Co-Trimoxazole (see appendix B)

Important of antibiogram are useful in detecting potential infectious disease outbreaks the use of such aggregate data on local or regional resistance trends is fundamental to discern differences and changes in patterns for appropriate selection of antimicrobials for rational use and epidemiological surveillance<sup>(16,17)</sup>

(Objectives): Prevalence of most microorganism in neonate

Develop antibiogram for NICU.





## **Chapter Two**

### **Literature Review**





# LITERATURE REVIEW

## 2.1 Infection in neonatal

Neonates are uniquely susceptible to infection. An immature immune system is coupled with exposure to the variety of maternal and environmental pathogens that can affect this population.<sup>(18)</sup>

Infants under the age of 3 months are rapidly building up their immunity by ramping up their production of immune cells and the creation of “memory” in their adaptive immune systems through various exposures to their environment. During this period, the child is very vulnerable to serious bacterial infections such as those caused by group B streptococcus (GBS) and *E. coli*.<sup>(19)</sup>

Common infectious diseases affecting children include bronchiolitis, pneumonia, urinary tract infection, sinusitis, skin infection, gastroenteritis, and acute otitis media.<sup>(20)</sup> **Whereas Common infectious diseases affecting in Neonatal sepsis and pneumonia Bloodstream infections (BSIs) are the most common HAIs in the NICU.**

They can occur in isolation or in association with urinary tract infections (UTIs) and meningitis. Endocarditis, osteomyelitis, pyogenic arthritis, ventilator associated pneumonia, peritonitis, conjunctivitis, and skin abscesses are important, less common HAIs.<sup>(18)</sup>

Whereas viral infection, Neonates, like older children and adults, are subject to viral infections acquired by horizontal routes, such as those due to influenza, rotavirus, and enteroviruses.

They also are at risk for viruses through routes that are unique to the perinatal setting in which mother-to-child transmission can occur transplacentally, during birth, or from breast milk. The ability of cytomegalovirus (CMV), herpes simplex virus (HSV), human immunodeficiency virus (HIV), and human T-lymphotrophic virus type 1 (HTLV-1) to establish chronic infection in the mother with persistence of infectious virus in blood, mucosa, or milk accounts for the role vertical transmission plays in their epidemiology and potential clinical impact.

Whether viruses that produce acute, self-limited infections in the mother are transmitted to the fetus or newborn depends on the timing of maternal infection in relation to gestation



and parturition. The clinical settings in which fetal and neonatal viral infections must be considered include pregnancy, the newborn nursery, and the evaluation of an ill newborn.<sup>(18)</sup>

## 2.2 Antimicrobial Resistance (AMR)

Occurs when bacteria, viruses, fungi and parasites change over time and no longer respond to medicines making infections harder to treat and increasing the risk of disease spread, severe illness and death.

### Mechanisms of Resistance .

1. Intrinsic resistance – Gram-negative bacteria have an outer membrane that makes them less permeable than gram-positive bacteria.
2. Adaptive resistance – Bacteria can adapt to their environment to survive. This occurs in the development of biofilms.
3. Acquired resistance – Most resistance that is of alarm is acquired resistance, with bacteria passing genetic material on plasmids.
  - a. Enzymes such as  $\beta$ -lactamases are passed on plasmids – Affected:  $\beta$ -Lactams.
  - b. Bacteria have reduced permeability or can use efflux pumps to push antibiotic out of the cell – Affected: Aminoglycosides, fluoroquinolones, macrolides, tetracycline.
  - c. Alteration in drug-binding target – Affected:  $\beta$ -Lactams, oxazolidinones, aminoglycosides, tetracycline, glycopeptides.
  - d. Alteration of the antibiotic – Affected: Fluoroquinolones, aminoglycosides, lincosamides
  - e. Bypass the effect of the antibiotic – Affected:  $\beta$ -Lactams, sulfamethoxazole/trimethoprim, glycopeptides <sup>(21)</sup> .

Antimicrobial resistance classified into:

### 2.2.1 Resistance in Gram-Negative Infections

#### Mechanisms of Resistance.

The common thread in most of the organism threats is the presence of  $\beta$ -lactam resistance.

- a)  $\beta$ -Lactams are the largest (and most prescribed) class of antimicrobials: penicillins, cephalosporins, monobactams, and carbapenems .
- b) Three principles of acquired  $\beta$ -lactam resistance:
  - i) Decreased outer membrane penetration – Porin loss and increased drug efflux pumps .
  - ii) Alteration in binding target.
  - iii) Production of  $\beta$ -lactamases .





#### Extended-Spectrum $\beta$ -Lactamases:

- (1) Ability to hydrolyze penicillins, third-generation cephalosporins, and monobactams. Do not hydrolyze cephamycins and carbapenems.
- (2) The three main families are TEM, SHV, and CTX-M. The most prevalent type is CTX-M, distributed worldwide and increasing in prevalence, found in *E. coli* and *K. pneumoniae*. Gene originally produced by bacteria *Kluyvera*.
- (3) Originally associated with hospital-acquired outbreaks; now found in community-acquired infections.
- (4) Associated with high mortality.

#### Carbapenems-Resistant Enterobacteriaceae

- i) Rise in prevalence is related to the selective pressure from increasing carbapenems use.
- ii) Carbapenems resistance in the United States is the combination of several mechanisms (e.g., ESBL plus poring loss), and these CRE do not constitute a carbapenems-producing Enterobacteriaceae (CPE). This distinction likely matters more epidemiologically than clinically because testing is not routine.
  - (a) True CPE likely has a very high carbapenem MIC, whereas CRE may have lower carbapenem MIC (though still resistant).
  - (b) CPE has been associated with higher mortality because it may represent a more virulent organism type.
- iii) Organisms typically have acquired resistance to many classes by several mechanisms, and many isolates show as extremely drug resistant.
- iv) Risk factors for poor outcomes include ICU admission, advanced age, requirements for vasopressors, mechanical ventilation, dialysis, non-urinary-sourced infections, and presence of colistin-resistant CRE. <sup>(22,23)</sup>

#### 2.2.2 resistance in Gram-Positive Infections

##### A. *S. aureus* Mechanisms of :

- (1) Resistance has evolved for *S. aureus* depending on antibiotic exposure over time, starting with penicillin once exposed.
  - (a) Penicillin resistance developed 1 year after introduction of penicillin.
  - (b) Methicillin resistance developed 2 years after introduction of methicillin.
- (2) MRSA
  - a. Penicillin resistance is caused by  $\beta$ -lactamase production (penicillinases). Methicillin is stable against hydrolysis by penicillinases.





b. Methicillin (and other  $\beta$ -lactam) resistance comes from alteration in the penicillin-binding protein (PBP)  $\beta$ -lactam-binding target to PBP2a, which is coded for by the *mecA* gene, carried on the staphylococcal cassette chromosome (SCC) SCCmec.

c. Theories regarding the origin of MRSA suggest that SCCmec was first transferred from coagulase-negative staphylococci.

(3) Vancomycin-resistant *S. aureus* (VRSA)

a. Occurrence is limited, with less than 20 incidents reported worldwide.

b. Transmission of VanA gene from *Enterococcus* to *S. aureus*. Despite VRE arising in the 1980s, transmission to *S. aureus* has not been commonplace

B. *Enterococcus*

Normal human flora in the gastrointestinal tract and a hardy, but nonvirulent microbe. Second to staphylococci in cause of hospital-acquired infection.

Capable of transmitting genes easily

Mechanisms of resistance

a. Ampicillin resistance

i. Most hospital-based *E. faecium* harbor an altered PBP5 that conveys ampicillin resistance.

ii. *Enterococcus faecalis* resistance is mediated by  $\beta$ -lactamase enzymes or alteration in PBP4.

b. Vancomycin resistance

More common in *E. faecium* than in *E. faecalis*

Alteration in the D-Ala-D-Ala binding site

(a) D-Ala-D-Lac is associated with high-level vancomycin resistance.

(b) D-Ala-D-Ser is associated with low-level vancomycin resistance <sup>(21)</sup>





## **Chapter Three**

### **Methodology**





## Methodology

### 3.1 Patients and Study Site, study design

The study was a retrospective study undertaken at USTH in Sana'a, Yemen. USTH is a private hospital 165-bed teaching hospital that trains medical students, nurses, and pharmacists and serves as a referral hospital for the country.

The collected data from bacterial isolates from Jan 2021 to March 2022 with susceptibility testing performed using diffuse disk by the microbiology laboratory from the Neonates intensive critical care (NICU) settings at USTH.

### 3.2 Lab Techniques

Specimens from blood, cerebrospinal fluid, sputum, urine, stool and swab were collected, processed, and analyzed in the microbiology laboratory according to the Kirby-Bauer method. The Kirby-Bauer test for antibiotic susceptibility (also called the *disc diffusion test*) is a standard that has been used for years.

### 3.3 Data Collection

The list of all isolates was collected from the system, then the sensitivity results for positive isolates were checked from the hospital records system. The data was entered into WHONET 5.6, a free Windows-based database software developed for the management of microbiology laboratory data. The data entered for each culture specimen included specimen number, sex, age category, department, specimen date, and organism. (See appendix B)

### 3.4 Data analysis

WHONET aggregated and analyzed the data. Getting Started-Setting up an analysis: %R, I, S and test measurements- Running the analysis and interpreting the Results-Transferring WHONET results to Excel and other software Susceptible Summary Isolate listing and summary (see Appendix C)

### 3.5 Antibigram Development

Aggregated data from WHONET produced susceptibility percentages for every organism. The research team reviewed these auto-generated susceptibilities.

We initially excluded organisms not commonly associated with a disease or with fewer than 30 isolates, given the potential for diminished accuracy.<sup>(24)</sup>





We then reviewed the list and chose to include clinically important organisms despite having fewer than 30 isolates with the notation that these results should be interpreted with caution based on the low number of isolates.

The antibiotics included in the antibiogram were narrowed to those commonly available at USTH. We developed one antibiogram for pediatric doctors with specific percentage details for the most common antibiotics, and antibiotic sensitivity represented as resistant (“R”), intermediate (“I”), and sensitive (“S”). A common, but not universal, practice is to define susceptible as 80% to 100% susceptible, intermediate as 60% to 79.9% susceptible, and resistant as 0–59.9% susceptible.

We used these ranges in this antibiogram. In the instance of intrinsic resistance of an organism to an antibiotic, this was labeled as “R” rather than providing the percentage susceptible.

### 3.6 Ethics Statement

The study was approved by EIU and USTH. because this was a retrospective study of de-identified specimens, no consent form was required. (see appendix D)





## **Chapter Four**

### **Result**





## Result

The number of screened patients were (257) with 324 isolated and out of them there were (133) patients with (148) positive isolation. The patients were screened from 2021/1/1 to 2022/3/31. The male represents 74% and female 26% (Table 1 gender numbers)

**Table 1 gender of isolated**

Sex	Frequency of isolated	(%)	Frequency of patients
M	109	74	97
F	39	26	36
Total	148	100	133

Table 2 showed that blood source represented the most isolation 133 (89.9%), while other sources (i.e. urine, swab, sputum, ....) were not very common source for isolation.

**Table 2 The source of isolations with their Frequency and Percent .**

Source	Frequency	Percent %
Blood	133	89.9%
Urine	4	2.7%
Swab	3	2%
Sputum	3	2%
Cerebrospinal fluid	2	1.4%
Other	2	1.4%
Stool	1	0.7%





#### 4.1 Isolated and number of bacterial species that included in the antibiogram .

The total number of isolated after removal of rare bacteria isolated was 138 (G + ve (94 (68.1%)), G -ve bacteria (44 (31.9%)). The results showed that G +ve bacteria was the most encounter isolated in USTH NICU, followed by G -ve bacteria.

**Table 3 bacterial species isolated and its number at NICU**

Bacterial species	No isolates	(%)	No patients	$X^2 / P$
Acinetobacter sp.	3	2.2	3	0.056/1
Burkholderia cepacia	9	6.5	8	
Enterobacter sp.	3	2.2	3	
Escherichia coli	6	4.3	6	
Klebsiella pneumoniae	5	3.6	5	
Klebsiella sp.	12	8.7	11	
Pseudomonas sp.	6	4.3	6	
Total of GN Isolates	44	32	42	

**Table 4 bacterial species isolated and its number at NICU**

Bacterial species	No isolates	(%)	No patients	$X^2 / P$
Staphylococcus aureus	6	4.3	6	1.734 /0.78445
Enterococcus sp.	6	4.3	6	
Staphylococcus hominis	4	2.9	4	
Staphylococcus coagulase negative	69	50	68	
Streptococcus sp.	9	6.5	7	
Total of GP Isolates	94	68	91	





#### 4.1.1 Isolated of gram positive of bacterial species

##### 4.1.1.1 staphylococcus coagulase negative: -

Staphylococcus was the most isolated G+ve 69 (50%) most antibiotic sensitive is linezolid and vancomycin as Table 5, The most

*Table 5 Staphylococcus, coagulase negative sensitivity*

Bacterial sppecies	Staphylococcus, coagulase negative		
AB	Total	No sensitive	Percent %
Amikacin	58	48	82.8
Azithromycin	54	10	18.5
Cefuroxime	56	23	41.1
Clindamycin	66	44	66.7
TMP/SMX	67	27	40.8
Erythromycin	69	10	14.5
Gentamicin	65	39	60
Linezolid	65	65	100
Penicillin	7	1	14.3
Tigecycline	4	4	100
Vancomycin	67	67	100



**4.1.1.2** Streptococcus spp with was the 2<sup>nd</sup> isolated G +ve9 (6.5%) with sensitive to linezolid and vancomycin as is in (Table 6)

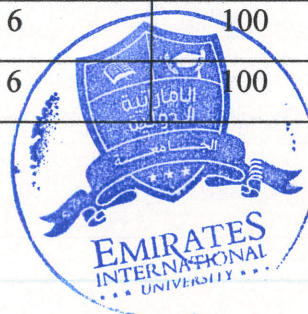
**Table 6 Streptococcus sp sensitivity**

Bacterial species	Streptococcus sp		
AB	Total	No sensitive	Percent %
Azithromycin	6	5	86.3
Cefuroxime	9	6	66.7
Clindamycin	9	8	88.9
Linezolid	7	7	100
Penicillin G	9	7	77.8
Vancomycin	9	9	100

**4.1.1.3** Staphylococcus aureus with 6(4.3%) isolate, the most sensitive was Linezolid, vancomycin and amikacin as is in (table 7)

**Table 7 straphylococcus aureus sensitivty**

Bacterial species	Staphylococcus aureus		
AB	Total	No sensitive	Percent %
Amikacin	5	4	80
Azithromycin	6	1	16.7
Cefuroxime	6	2	33.3
Clindamycin	6	4	66.7
TMP/SMX	6	2	33.3
Erythromycin	6	1	16.7
Gentamicin	6	4	66.7
Linezolid	6	6	100
Vancomycin	6	6	100





4.1.1.4 Enterococcus sp is with 6 (4.3%) isolate, the most sensitive was linezolid and vancomycin as is in (table8)

**Table 8 Enterococcus sp. sensitive**

Bacterial species	Enterococcus sp.		
AB	Total	No sensitive	Percent %
TMP/SMX	6	0	0
Erythromycin	4	2	50
Linezolid	6	6	100
Penicillin G	6	2	33.3
Vancomycin	6	6	100

4.1.1.5 Staphylococcus hominis which is the less bacterial in gram +ve with 4(2.9%) with the most sensitive is linezolid, vancomycin and tigecycline as in (table9)

**Table 9 Staphylococcus hominis sensitive**

Bacterial species	Staphylococcus Hominis		
AB	Total	No sensitive	Percent %
Clindamycin	4	1	25
TMP/SMX	4	3	75
Erythromycin	4	0	0
Gentamicin	4	2	50
Linezolid	4	4	100
Tigecycline	4	4	100
Vancomycin	4	4	100





#### 4.1.2. Overall susceptibility of Gram-Positive Organisms

The above showed that There were (69 (50%)), Staphylococcus species, coagulase negative (CoNS) isolates, 9(6.5%) Streptococcus spp, 6(4.3%) S. aureus isolates, 6(4.3%) Enterococcus isolates, and 4(2.9%) Staphylococcus hominis. S had decreased susceptibility to Erythromycin 69(14.5%), while maintaining susceptibility to vancomycin, linezolid, while other antibiotics such as trimethoprim and cephalosporin were resistance to most gram + bacteria .

**Table 10 antibiogram of gram positive**

Organism	Orga nism	Amikacin n(%)	Azithromycin %S	Cefepime %S	Cefixime %S	Cefoperazone %S	Cefotaxime %S	Ceftazidime %S	Cefuroxime %S	Clindamycin %S	Colistin %S	Trimethoprim/Sulfameth oxazole %S	Erythromycin %S	Gentamicin %S	Imipenem %S	Linezolid 10 %S	Penicillin G 1 unit %S	Piperacillin/Tazobacta m 100/10 %S	Tigecycline %S	Vancomycin 5 %S	Polymyxin B %S	Meropenem %S	Ceftioxone %S
Staphylococcus aureus	6	S	R						R	I		R	R	I		S				S			
Staphylococcus, coagulase negative	69	S	R	R					R	I		R	R	I		S	R		S	S		S	
Staphylococcus hominis	4									R		I	R	R		S			S	S			
Streptococcus sp.	9		S						I	S		I	S			S	I			S			
Enterococcus sp.	6											R	R			S	R			S			

Colors green = sensitive (80-100%) yellow = intermittent (60-79.9%) red = resistance (0-59.9%)



### 4.1.3 Gram negative bacterial species

4.1.3.1 The number of isolated bacteria of *Acinetobacter* spp was 3(2.2%) and the most sensitive antibiotic for it were Colistin Imipenem... as in (table 11)

**Table 10 *Acinetobacter* sensitive**

Bacterial species	Acinetobacter sp		
AB	Total	No sensitive	Percent %
Amikacin	3	1	33.3
Cefepime	2	0	0
Cefotaxime	3	0	0
Ceftazidime	3	0	0
Colistin	2	2	100
TMP/SMX	2	1	50
Gentamicin	2	0	0
Imipenem	3	3	100
Piperacillin/Tazobactam	3	3	100
Polymyxin	1	1	100
Meropenem	3	3	100
` Ceftriaxone	2	0	0

4.1.3.2 the number of isolated bacteria of *Escherichia coli* was 6(4.3%), the most sensitive Amikacin, Gentamicin...etc. as in (table 12)

**Table 11 *Escherichia coli* sensitive**

Bacterial species	Escherichia coli		
AB	Total	No sensitive	Percent %
Amikacin	5	5	100
Cefepime	6	1	16.6
Cefixime	2	0	0
Cefoperazon	3	0	0
Cefotaxime	3	1	33.3
Ceftazidime	6	1	16.6



Cefuroxime	4	0	0
TMP/SMX	5	1	20
Gentamicin	6	6	100
Imipenem	5	5	100
Piperacillin/Tazobactam	4	2	50
Meropenem	6	6	100
Ceftriaxone	6	1	16.7

4.1.3.3 The number of isolated bacteria of Enterobacter spp is bacteria from was 3(2.2%),  
The most antibiotic sensitive is in (table 13)

**Table 12 Enterobacter sp sensitive**

Bacterial species	Enterobacter spp		
	Total	No sensitive	Percent %
AB			
Amikacin	2	1	50
Cefepime	2	0	0
Cefixime	1	0	0
Cefotaxime	2	1	50
Ceftazidime	2	0	0
Cefuroxime	2	0	0
TMP/SMX	3	2	66.7
Gentamicin	2	1	50
Imipenem	3	2	66.7
Penicillin G	1	1	100
Piperacillin/Tazobactam	2	1	50
Meropenem	2	1	50
Ceftriaxone	2	0	0



4.1.3.4 *Klebsiella* spp is the most gram negative bacterial found in this study with number of isolate 12(8.7%), the most sensitive antibiotic colistin and polymyxin as in (table 14)

**Table 14 *Klebsiella* sp sensitive**

Bacterial species	<i>Klebsiella</i> sp		
AB	Total	No Sensitive	Percent %
Amikacin	12	7	58.3
Cefepime	12	1	8.3
Cefixime	3	0	0
Cefoperazon	6	0	0
Cefotaxime	4	1	25
Ceftazidime	12	0	0
Cefuroxime	6	0	0
Colistin	1	1	100
TMP/SMX	11	7	63.6
Gentamicin	11	4	36.4
Imipenem	7	6	85.7
Piperacillin/Tazobactam	7	4	57.1
Polymyxin B	1	1	100
Meropenem	12	7	58.3
Ceftriaxone	11	0	0

4.1.3.5 the number of isolated bacteria of *Klebsiella pneumonia* is was 5(3.6%) isolate, the most sensitive antibiotic is in (table 15)

**Table 15 *Klebsiella pneumonia* sensitive**

Bacterial species	<i>Klebsiella pneumoniae</i>		
AB	Total	No sensitive	Percent %
Amikacin	5	5	100
Cefepime	5	1	20
Ceftazidime	5	1	20
TMP/SMX	5	4	80
Meropenem	5	4	80
Ceftriaxone	5	1	20



4.1.3.6 The number of isolated bacteria of *Pseudomonas* spp was 6(4.3%) isolates, the most sensitive antibiotic is in (table 16)

**Table 13 *Pseudomonas* sp sensitive**

Bacterial species	Pseudomonas sp		
AB	Total	No sensitive	Percent %
Amikacin	6	3	50
Cefepime	5	1	20
Cefoperazon	1	0	0
Ceftazidime	6	1	16.7
Cefuroxime	1	0	0
Gentamicin	6	3	50
Meropenem	6	6	100
Ceftriaxone	5	2	40

4.1.3.7 *Burkholderia cepacia* is 2<sup>nd</sup> gram negative Bactria in this study with (9(6.5%)) isolated and the most sensitive antibiotic is in (table 17)

**Table 14 *Burkholderia cepacia* sensitive**

Bacterial species	Burkholderia cepacia		
AB	Total	No sensitive	Percent %
Ceftazidime	9	4	44.4
TMP/SMX	9	6	66.7
Meropenem	8	8	100





#### 4.1.4 The above showed that Gram-Negative Bacteria

Klebsiella spp and Burkholderia cepacia were the most isolated Gram negative (GN) bacteria, with 12(8.7%) and 9(6.5%) specimens, they were highly resistant to all antibiotics except Imipenem, Polymyxin B and Colistin for Klebsiella spp and Meropenem, Trimethoprim/Sulfamethoxazole for Burkholderia cepacia. Klebsiella spp had susceptibilities of Piperacillin/Tazobactam 7(57.1%), Cefotaxime 4(25%) and Gentamicin 11(36.4%). The high resistance to 3rd. generation cephalosporin indicated high rates of beta-lactamase production including ESBLs

**Table 15 Antibigram for gram negative**

Organism	Org	Amikacin n(%)	Azithromycin %S	Cefepime %S	Cefixime %S	Cefoperazone %S	Cefotaxime %S	Ceftazidime %S	Cefuroxime %S	Clindamycin %S	Colistin %S	Trimethoprim/Sulfamethoxazole	Erythromycin %S	Gentamicin %S	Imipenem %S	Linezolid 10 %S	Penicillin G 1 unit %S	Piperacillin/Tazobactam 100/10	Tigecycline %S	Vancomycin 5 %S	Polymyxin B %S	Meropenem %S	Ceftriaxone %S
Acinetobacter sp.	3	R		R			R	R			S	R		r	S			S			S	S	R
Escherichia coli	6	S		R	R	R	R	R	R			R		S	S			R				S	R
Enterobacter sp.	3	R		R	R		R	R	R			I		R	I		S	R				R	R
Klebsiella sp.	12	R		R	R	R	R	R	R		S	I		R	S			R			S	R	R
Klebsiella pneumoniae	5	S		R				R				R		S								S	R
Burkholderia cepacia	9							R				I										S	
Pseudomonas sp.	6	R		R		R		R	R					R	S			S				S	R

Colors green = sensitive (80-100%), yellow = intermittent (60-79.9%), red = resistance (0-59).



## **Chapter Five**

### **Discussion**





## Discussion

In the study, were analyzed neonate bacterial isolates from USTH at NICU and most important findings were as following: (1) G +ve were the most isolated bacteria among neonate patients; (2) the most sensitive antibiotics for gram +ve were linezolid and vancomycin; (3) The most isolated g -ve bacteria were resistance to most cephalosporin generations and sensitive mostly for carbapenem antibiotics.

In this study the gram positive was the most isolated bacteria in USTH hospital institution the most common isolate *Staphylococcus coagulase negative* 69 (50%). however, in other studies that were conducted at NICU form Zambia, (25) and in Yemen; was in Sana'a (26).

They found the gram negative was the most prominent bacteria. The high percentage of gram positive in our institution may be due to contaminated but we cannot confirm that in our study as retrospective study design. Other factors may be due to different locations, populations, and clinical situation. For the gram-positive bacteria, the most sensitive antibiotic vancomycin and linezolid and resistance to cephalosporin, Erythromycin and TMP/SMX. Similar to our finding, studies that was conducted in Yemen, and Zambia found that TMP/SMX and erythromycin were resistance as well. The most sensitive antibiotics in our study and mention studies were vancomycin and linezolid. *Staphylococcus* resistance to TMP/SMX and erythromycin indicates the isolated *Staphylococcus* was MRSA. this is indicted for staph auras the MRSA the most commonly encounter bacteria and this is may be duo to result from the overuse of antibiotics.

Our findings of high resistance to Erythromycin and TMP/SMX are consistent with other studies. These findings are concerning because TMP/SMX is commonly used for empiric treatment, particularly respiratory and gastrointestinal infections.

The gram negative isolates showed high level of sensitivity to carbapenem and high resistance to cephalosporin which was consistent with other studies that were conducted in Zambia, and this is maybe indicate of ESBL for these isolated.





## 5.2 Limitation

This study had some limitations. First, this retrospective study is based on the data collected from laboratory records which lack information about the neonates' hospitalization date, clinical information, and treatment outcome. Therefore, we were not able to classify infections as community acquired or hospital acquired infection. Similarly, we could not determine whether the antibiotic resistance was primary or secondary resistance. Moreover, data on the clinical information and treatment outcome of the neonates were not included in this study. Second, this study was conducted only at a single hospital; therefore, the antibiotic resistance patterns observed in our study might not generalize the situation in the country.





## **Chapter Six**

### **Conclusions and Recommendation**





## 6.1 Conclusions

The resistance for antimicrobial gram positive and gram negative still high in neonate as only vancomycin and linezolid was most sensitive for the gram positive and carbapenem and Colistin only the sensitive antibiotic for gram negative

## 6.2 Recommendation

Antibiogram is necessary for neonate as national level not local level prospective Should be restriction use of unnecessary antibiotic and awareness healthcare and student about resistance because there is high rate of resistance among neonate

Focus the infection control to limited the separate the most resistance bacteria





## References

- 1) Black, R.E.; Cousens, S.; Johnson, H.L.; Lawn, J.E.; Rudan, I.; Bassani, D.G.; Jha, P.; Campbell, H.; Walker, C.F.; Cibulskis, R.; et al. Global, regional, and national causes of child mortality in 2008: A systematic analysis. *Lancet* 2010, 375, 1969–1987.
- 2) Basha, S.; Surendran, N.; Pichichero, M. Immune responses in neonates. *Expert Rev. Clin. Immunol.* 2014, 10, 1171–1184.
- 3) Sass, L.; Karlowicz, M.G. Healthcare-Associated Infections in the Neonate. *Princ. Pract. Pediatr. Infect. Dis.* 2018, 3, 560–566.
- 4) Cortese, Francesca, et al. "Early and late infections in newborns: where do we stand? A review." *Pediatrics & Neonatology* 57.4 (2016): 265-273.
- 5) Sharif MR, Alizargar J, Sharif A. Antimicrobial resistance among Gram-negative bacteria isolated from different samples of patients admitted to a university hospital in Kashan, Iran. *Adv Biol Res* 2013; 7: 199–202.
- 6) Patzer JA, Dzierzanowska D, Turner PJ. Trends in antimicrobial susceptibility of Gram-negative isolates from a paediatric intensive care unit in Warsaw: results from the MYSTIC programme (1997-2007). *J Antimicrob Chemother* 2008; 62: 369–75.
- 7) Centers for Disease Control and Prevention. Antibiotic Resistance Threats in the United States, 2019, 2019.
- 8) WHO. Global action plan on antimicrobial resistance. *World Heal Organ.* 2017;1–28.
- 9) Vazouras, K.; Velali, K.; Tassiou, I.; Anastasiou-Katsiardani, A.; Athanasopoulou, K.; Barbouni, A.; Jackson, C.; Folgori, L.; Zaoutis, T.; Basmaci, R.; et al. Antibiotic treatment and antimicrobial resistance in children with urinary tract infections. *J. Glob. Antimicrob. Resist.* 2020, 20, 4–10.
- 10) World Health Organization. Antimicrobial Resistance Global Report on Surveillance; World Health Organization: Lyon, France, 2014. Available online:
- 11) Wangai, F.K.; Masika, M.M.; Lule, G.N.; Karari, E.M.; Maritim, M.C.; Jaoko, W.G.; Museve, B.; Kuria, A. Bridging antimicrobial resistance knowledge gaps: The East African perspective on a global problem. *PLoS ONE* 2019;14, e0212131.





- 12) Carroll, M.; Rangaiahagari, A.; Musabeyezu, E.; Singer, D.; Ogbuagu, O. Five-Year Antimicrobial Susceptibility Trends Among Bacterial Isolates from a Tertiary Health-Care Facility in Kigali, Rwanda. *Am. J. Trop. Med. Hyg.* 2016, 95, 1277–1283.
- 13) Pengsaa K, Lumbiganon P, Taksaphan S, Pairojkul S, Sookpranee T, Kosuwon P, et al. Risk factors for neonatal Klebsiella septicemia in Srinagarind Hospital. *Southeast Asian J Trop Med Public Health* 1996; 27:102–6.
- 14) Mohammed K, Abdulrahman Z, Dalal A, Mahmoud A. Antibiotic Resistance Trends of Gram-negative Bacteria. ORIGINAL ARTICLE.2018
- 15) Wafa F. S. Badulla, Mohammed A, Mohamed I. Antimicrobial Resistance Profiles for Different Isolates in Aden, Yemen. *Hindawi*.2020;5
- 16) WHO. Organization World Health. Global Strategy for Containment of Antimicrobial Resistance. *World Heal Organ.* 2001;105.
- 17) Avdic E, Carroll KC et al. The role of the microbiology laboratory in antimicrobial stewardship programs. *Infect Dis Clin North Am.* 2014; 28:215–35.
- 18) Sarah S, Charles G; Principles and Practice of Pediatric Infectious Diseases 2018 ELSEVIER 5<sup>th</sup> edition.
- 19) Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proceedings. Biological sciences.*
- 20) Zeimet A, McBride DR, Basilan R, Roland WE, McCrary D, Hoonmo K. Infectious diseases. *Textbook of Family Medicine.*
- 21) Katherine L. Infectious board of clinical pharmacy antimicrobial resistance 2018 1.3-33
- 22) Infectious Disease Society of American for melty drugs resistance 2022.
- 23) European society of infectious disease and multy drug resistance 2022
- 24) World Health Organization. Antimicrobial Stewardship Programmes in Health-Care Facilities in Low- and Middle-Income Countries: A WHO Practical Toolkit; World Health Organization: Lyon, France, 2019.
- 25) Brenna M, Alexandra L, Cassidy C. Antibigram Development in the Setting of a High Frequency of Multi-Drug Resistant Organisms *MDPI* 2021.
- 26) Adeeb S, Ibrahim S, Mohmmmed A. Neonatal sepsis in Sana'a city, Yemen: a predominance of Burkholderia cepacia; *BMC*;2021.



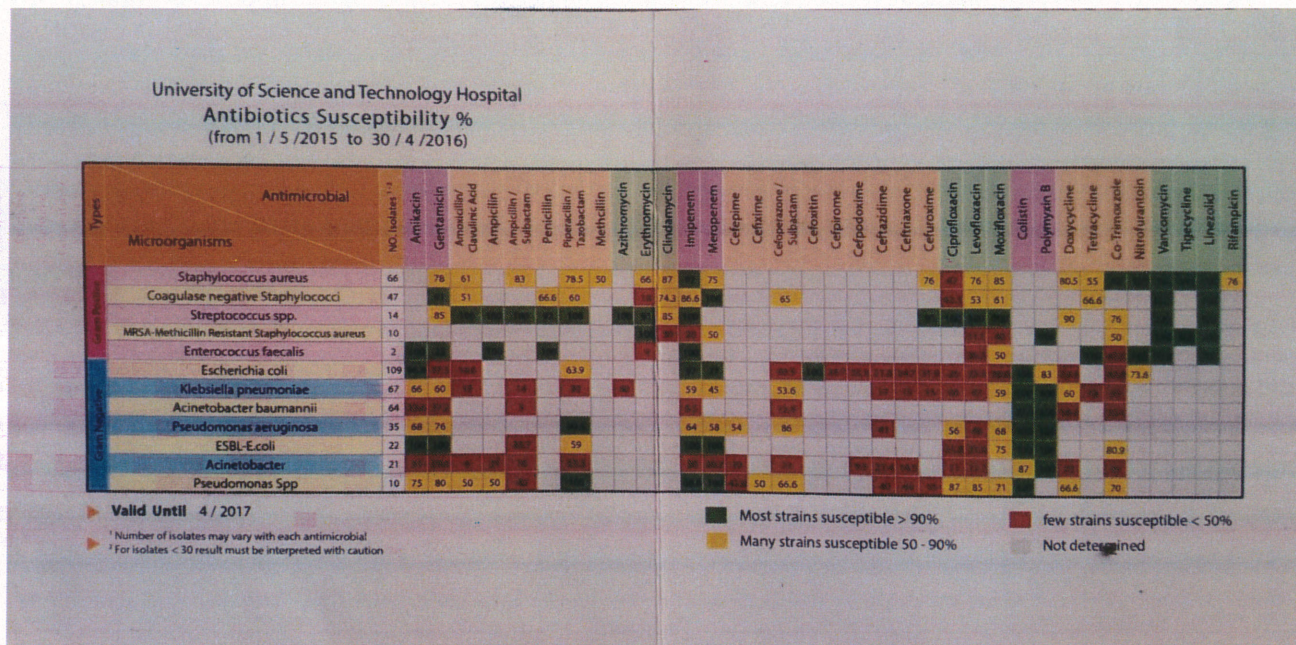


## APPENDICES





## Appendix A Pervious Work





## Appendix B Data collected

**Data fields** [X]

Your data fields appear below.  
Make any necessary changes.  
If you want to add or remove fields, select 'Modify list'.

neonatal university of science and technology hospital

Country	Laboratory	Identification number	Last name	First name	Sex	Age	Age category	Institution	Department	Location type	Specimen number	Specimen date	Specimen type	Specimen type (Numeric)	Isolate number	Organism	Organism type	Beta-lactamase	ESBL	Carbapenemase

Number of fields = 24

**Code list**

File origin: [Unicode] Character set: [Unicode (UTF-8)]

**Serotype**

Description	Serotype
Name	SEROTYPE
Type	Text
Length	40

**Code list**

SEROTYPE.txt

**Data entry**

Section: [Microbiology]

☒ Human  
☒ Animal  
☒ Food

☐ Isolate listing

**OK**



## Appendix C Data analysis

Data analysis: neonatal university of science and technology hospital

Analysis type Options One per patient

Organisms Isolates

Data files Output Screen

Macros Begin analysis Exit

Laboratory

Country code	Laboratory code	Laboratory name
WHO	AGI	WHO AGISAR Sample data
WHO	GLS	WHO GLASS Demonstration
WHO	TST	WHO Test Laboratory
YEM	NUSTH	neonatal university of science and technology hospital

New laboratory

Open laboratory

Modify laboratory

Copy laboratory

Delete laboratory

Language and dates

Select fonts

Browse C:\WHONET\





Analysis selection - %RIS and test measurements

Isolate listing and summary | %RIS and test measurements | Scatterplot | Resistance profiles | Isolate alerts | Cluster alerts

Report format

☒ 1. %RIS and test measurements

☒ Tables

☒ Graphs

☐ 2. Summary

☐ Tables

☐ Graphs

Antibiotics

☒ All antibiotics ☐ Select antibiotics

Options

Percentage or number

☒ Percentage of isolates

☐ Number of isolates

Measurement ranges

Disk diffusion 6 - 35 mm

MIC and Etest .002 - 256 ug/ml

Histograms

☒ Breakpoints

☐ Quality control

ATCC 25922 (eco)

☐ MIC panels

ASCAM Campylobacter

☐ Display the histogram's legend.

☐ Use new WHONET breakpoint tables

Organisms

Select the organisms that you would like to include in the analysis.  
Make your selections by double-clicking or by typing the codes and pressing <Enter> after each one.

WHONET organism list

Code ALL

☐ Extended list ☒ Organism groups

ALL All organisms  
GM+ Gram positive organisms  
GM- Gram negative organisms  
ANA Anaerobes  
MYC Mycobacteria  
FUN Fungi  
PAR Parasites  
OTB Other bacteria  
OTH Other organisms  
EBC All Enterobacteriaceae  
NFR All non-fermenting gram negative rods  
AC- Acinetobacter sp.  
AEC Aerococcus sp.  
AER Aeromonas sp.  
BCS Bacillus sp.  
BAC Bacteroides sp.  
BUK Burkholderia sp.  
CAM Campylobacter sp.  
CAN Candida sp.  
CI- Citrobacter sp.  
CDF Clostridium difficile

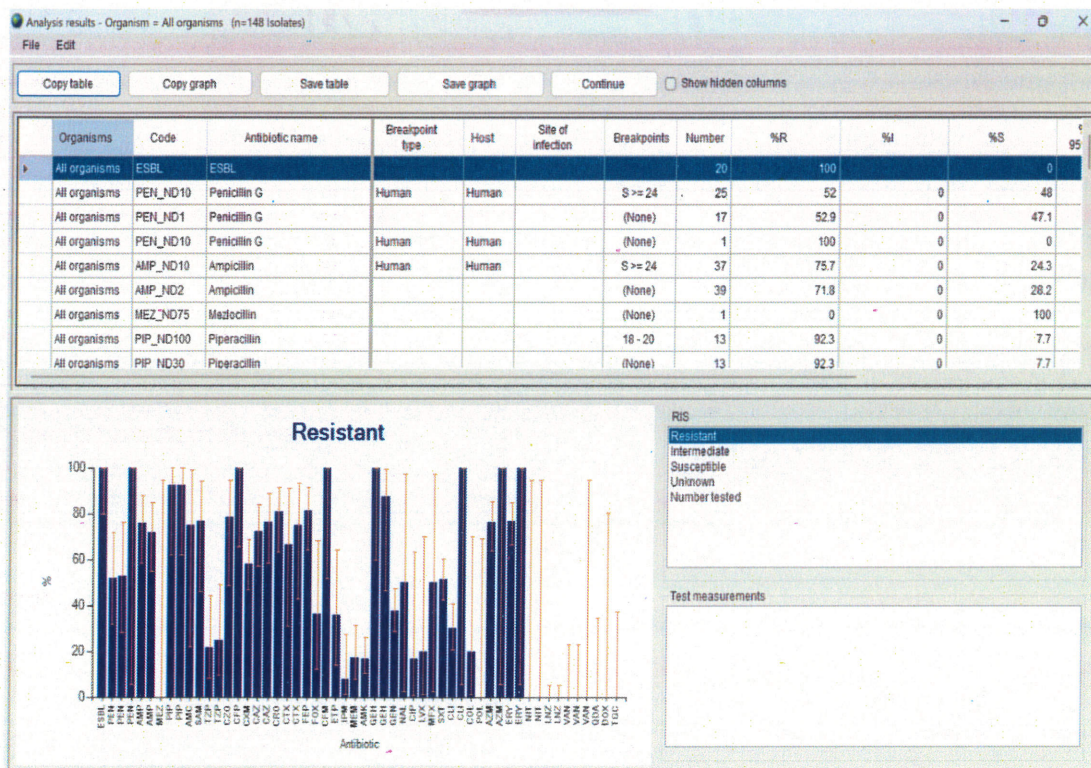
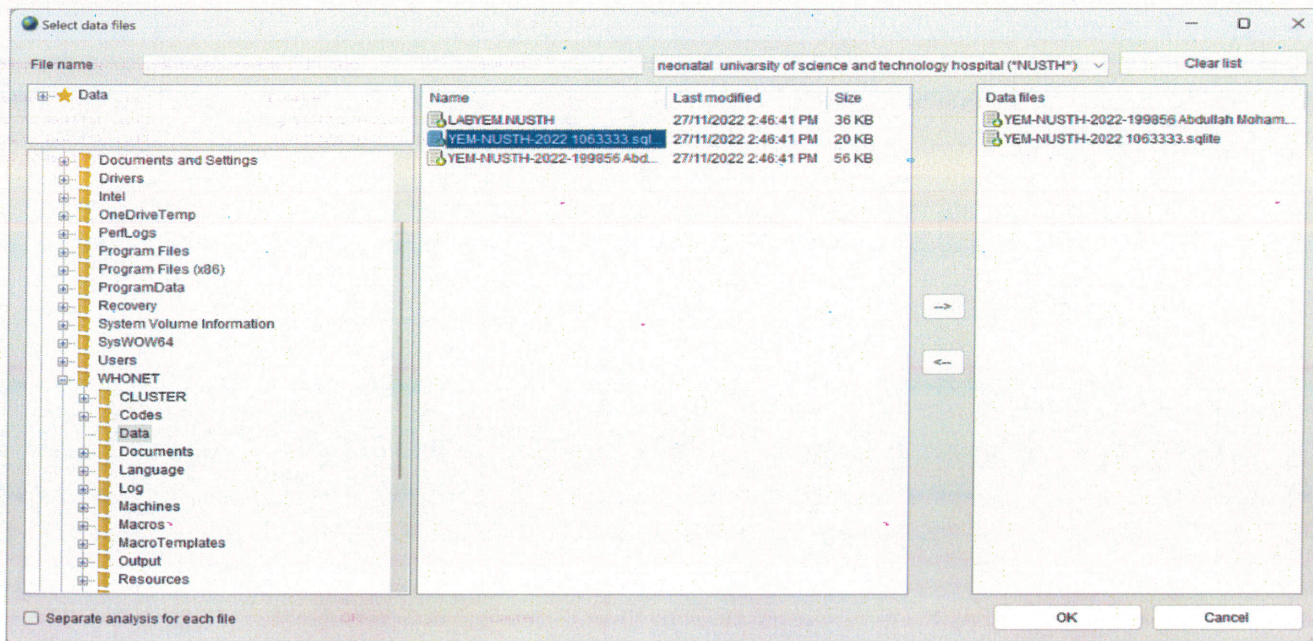
Analysis organism list

☐ Analyze as one organism

ALL All organisms

Search







## Appendix D Premission letter of EIU

Republic of Yemen  
Emirates International University  
Sana'a

الجمهورية اليمنية  
الجامعة الإماراتية الدولية  
صنعاء

المختبر / مستشفى جامعة العلوم والتكنولوجيا  
تحت إشراف دكتور

الموضوع / جمع بيانات بحث تخرج  
عندكم الجامعة الإماراتية الدولية أطيب التحايا متمنين لكم دوام التوفيق والنجاح في أعمالكم  
وبالإشارة إلى الموضوع أعلاه، نحيطكم علماً أن الطلاب التالية أسمائهم

- 1- ايمن الحيشي
- 2- مسعود رسام
- 3- عمرو ناشر
- 4- نعتان عبدالمجيد
- 5- مجدي سيف
- 6- علاء الدين الصلوي
- 7- خالد العديني
- 8- محمد القزيلي
- 9- لول الذبحاتي
- 10- غاده صلاح
- 11- يسرى المشملي

من طلاب الجامعة الإماراتية الدولية المقيدين بالمستوى السادس في كلية الطب والعلوم الصحية قسم  
(Pharm-D)  
Specific Antibiotogram Development for ( Neonatal Intensive Care Unit at Referral Healthcare Center in Sanaa Yemen)

فترجو منكم التكرم والتعاون مع الطلاب مساهمة منكم مع الجامعة بربط الجانب العلمي بالجانب العملي  
هذا، وقد حررت هذا الرسالة بناءً على طلب الطلاب ولاتتحمل الجامعة أي مسؤولية تجاه الغير  
وتقبلوا خالص الشكر والتقدير ...

القائم بأعمال مدير شؤون الطلاب  
المؤيد الزواحي

صنعاء - حدة شارع بيروت/ تلخون: +9671415929 فاكس: +9671432222, Hada Beirut Street, Tel: +9671415929





## الخلاصة

### خلفية

مناعة حديثي الولادة منخفضة للغاية ويمكن أن تؤدي العدوى فيما بينها إلى عواقب تهدد الحياة. تعتبر مقاومة المضادات الحيوية واحدة من أكثر المجالات تحدياً في علاج الأمراض المعدية في جميع أنحاء العالم ويمكن أن تؤدي إلى فشل العلاج بين حديثي الولادة.

### الهدف

كان الهدف العام من هذه الدراسة هو تطوير مضاد حيوي خاص بحديثي الولادة في مستشفى جامعة العلوم والتكنولوجيا، صنعاء، اليمن.

### المنهجية

هذه دراسة بأثر رجعي، تم جمع البيانات من قاعدة بيانات مستشفى جامعة العلوم والتكنولوجيا وتم استخدام برنامج WOHNET لتحليل وتطوير مخطط المضادات الحيوية في مستشفى جامعة العلوم والتكنولوجيا

### نتيجة

كان البكتيريا موجبة الجرام هي الأكثر شيوعاً من البكتيريا التي تواجهها بين حديثي الولادة، تليها البكتيريا سالبة الجرام، من البكتيريا موجبة الجرام كانت المكورات العنقودية السلبية والمكورات العقدية هي الأكثر شيوعاً، وكانت الكليسيلا والبروكولديريا هي الأكثر شيوعاً في البكتيريا موجبة الجرام، وكان المضاد الحيوي الأكثر حساسية بالنسبة للبكتيريا موجبة الجرام هما الفانكوميسين والينزولاييد، في حين أن الكاربابينيم والكوليستين في البكتيريا سالبة الجرام

### الاستنتاج

المقاومة لعامل مضاد الميكروبات كانت عالية في حديثي الولادة وتشمل البكتيريا موجبة وسالبة الجرام.